

# Genetic structure and gene flow in *Beta vulgaris* subspecies *maritima* along the Atlantic coast of France

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**Abstract** Locating and quantifying genetic variation within crop wild relatives is an ongoing activity of gene banks tasked with *ex situ* conservation. Without detailed information about the population genetics of a species, geography often serves as a reasonable proxy for differentiation. With this in mind, this paper examines the genetic diversity and differentiation of *Beta vulgaris* subsp. *maritima* accessions collected along a well-studied latitudinal gradient along the French Atlantic coast of France as well as Corsica, for use as representative genotypes from the Mediterranean basin. The aim of this work is to identify the scale and magnitude of differentiation and diversity in this set of accessions, using both molecular and quantitative traits. We assessed clinal variation and admixture in genetic and morphometric data along this latitudinal gradient. Results from this study revealed a complex

pattern of recent gene flow and immigration on a historical biogeographic structure. Our data suggest that a presumed latitudinal cline is in reality an admixed mosaic of genotypes.

**Keywords** *Beta vulgaris* · Biogeography · Crop wild relative · Gene flow · Genetic diversity · Genetic structure

## Introduction

Variation in wild relatives of crop species is a critical source of adaptive variation useful to plant breeding (Harlan 1976; Tanksley and McCouch 1997; Gur and Zamir 2004; Maxted et al. 2006; McCouch et al. 2012). Sea beet (*Beta vulgaris* L. subsp. *maritima* (L.) Arcang. —*B. vulgaris* ssp. *maritima*—henceforth *Beta maritima*) is estimated to be the wild progenitor of cultivated beet (de Bock 1986; Letschert et al. 1994; Santoni and Bervillè 1995; Biancardi et al. 2012). This crop wild relative has become the major source of genetic diversity and a reservoir of genetic traits useful in improvement of cultivated beet, especially sugar beet (Frese et al. 2001; Panella and Lewellen 2007; Biancardi et al. 2012).

The sea beet populations along the French coast have been well studied by researchers focusing on the genetics of life history evolution and phenology (Van Dijk and Boudry 1992; Van Dijk et al. 1997; Hautekèete et al. 2002a, b; Boudry et al. 2002; Van

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Dijk 2009; Wagmann et al. 2012). These studies have described a gradient of variation that effects the transition from an annual to a biennial life history. The observed clinal pattern may be a consequence of post glacial biogeographic processes along the French Atlantic coast; the Mediterranean Basin was a refuge in the last ice-age for populations of *B. maritima* (Villain 2007; Villain et al. 2009).

Locating and quantifying the variation within crop wild relatives is an ongoing activity of gene banks that draws on methodology and analysis from evolutionary biology, ecological genetics and landscape genetics. Current and historical interactions between genetic drift, mutation and selection configure natural variation across a species range (reviewed by Holsinger and Weir 2009; Merimans and Hedrick 2011). These dynamics play out at different geographic scales but have the net effect of influencing gene flow and eventual admixture. A well-curated *ex situ* collection has a structure that can confirm genetic identity, identify duplicates or hybrids, estimate under-sampling of genetic diversity (i.e., gaps) and, ultimately, guide future management decisions about efficient sampling protocols and collection size (Lockwood et al. 2007; Maxted et al. 2008; Ramirez-Villegas et al. 2010; Reeves et al. 2012).

Genetic structure can be estimated in crop wild relatives using a number of complimentary methods that include assignment tests and other population genetic and phylogenetic approaches, which exploit variation measured through molecular markers. The goal of these analyses is to identify the range of gene dispersal among sampling locations and to identify major genetic lineages that make up the collection. Estimating gene dispersal patterns from multi-locus data has shifted from indirect estimates based on  $F_{st}$  (Slatkin and Barton 1989; Whitlock and McCauley 1999) to new statistical methods in population genetics that have been developed to better identify how genetic variation is structured across the landscape (Pritchard et al. 2000; Manel et al. 2003, 2005; Huelsenbeck and Andolfatto 2007). These methods are able to help reconcile cryptic patterns in genetic structure that may be due to recent migration events with much older biogeographic patterns (Wilson and Rannala 2003).

In this study we examine accessions of *B. maritima* collected along a latitudinal transect extending up the French Atlantic coast of France. In addition we included in our analysis a set of accessions along the

coast of Corsica for use as representative genotypes from the Mediterranean basin. The aim of this work is to identify the scale and magnitude of differentiation and diversity in this set of accessions, using both molecular and quantitative traits. In particular we are interested in assessing how genetic and morphometric data estimate clinal variation and admixture along this latitudinal gradient.

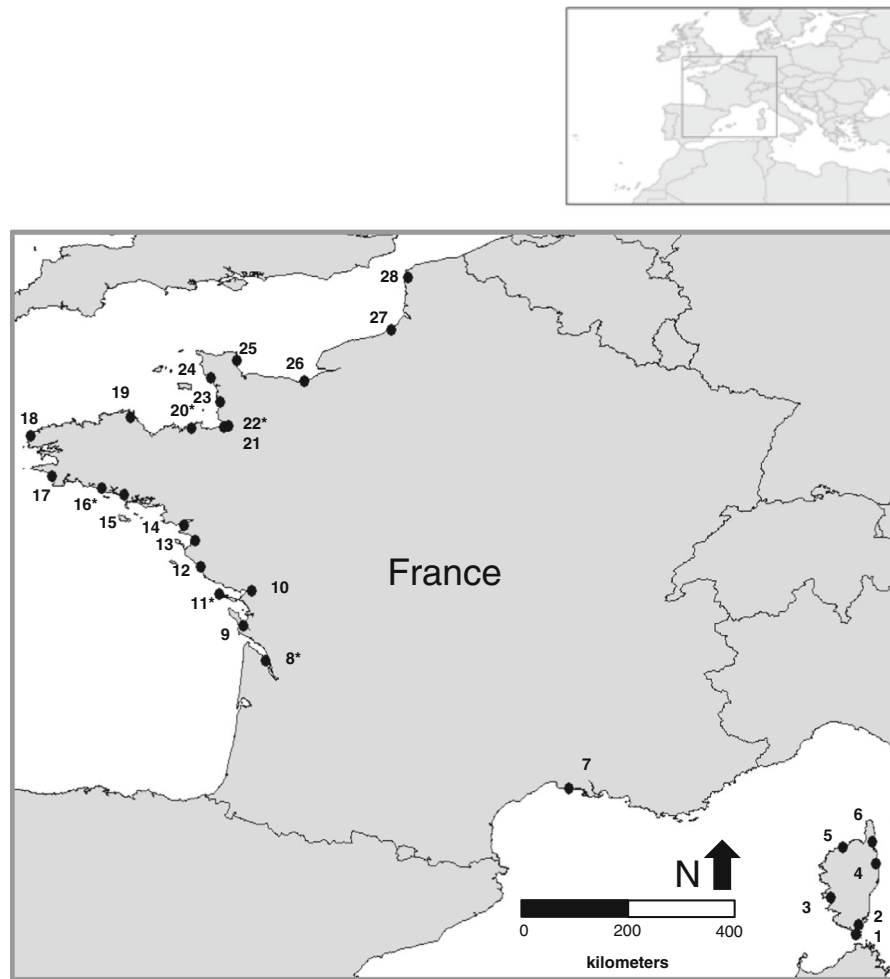
## Materials and methods

### Plant materials

Accessions used in this study were collected during two USDA-ARS sponsored plant exploration trips to Corsica (1985) and the French Atlantic coast of France (1989) (Fig. 1). Accessions were collected as bulked seed from 28 coastal sites that supported populations of 50–500 individuals (Table 1). These seeds from the original field collected accessions were evaluated in Crops Research Laboratory—Fort Collins Research Farm in Fort Collins, CO, at the USDA-ARS. The farm is located at about 40.672 N and 104.982 W and has a day length at the summer solstice of about 15 h and 5 min. The soil at this site is a Garrett loam, 0–1 % slope, pH 7.8, and was disked, roller harrowed and leveled in May prior to planting. Seeds were planted on June 1 and furrow irrigated as needed. All weeding was done by hand to avoid any herbicide phytotoxicity. The common garden trial used a randomized complete-block design with three, one-row plot replications for each of 28 accessions. Each of these replicated single row plots were 4 m long and were spaced 0.75 m apart. Each accession had 9–15 plants sown in each of the three plots. Plants were counted and numbered within each plot as they emerged. From each accession, 12 plants were chosen at random (4 from each plot) for phenotypic characterization and scored at appropriate times during the growing season. Leaf tissue also was collected from the same individuals for use in DNA extraction and marker analysis.

### Genotypic data

Genomic DNA was extracted from leaf tissue of four plants from each of the three replicate plots using Qiagen DNeasy 96 Plant Kits. With few exceptions, a total of 12 individuals for each of the 28 accessions were



**Fig. 1** Map of sampling locations for this study. Detailed descriptions given in Table 1

genotyped at 13 microsatellite (SSR) loci (Table 2) using a Licor 4200 DNA Sequencer (Licor, Lincoln, NE). PCR was carried out in 15  $\mu$ l total volume as previously described (Volk et al. 2005). Alleles were called manually using Licor Saga GT software. Duplicate samples from 12 individuals were extracted and genotyped to estimate scoring error rates. To improve scoring accuracy, loci were amplified and scored twice for all samples. Alleles from replicate amplifications were examined, and when alleles for replicates were not identical, data were entered as “missing”. Genotypic data was used if no more than one locus was missing.

#### Phenotypic data

We measured five foliar traits, collected from three mature leaves from each sampled individual. Leaf

length measured as the distance from leaf tip to the distal terminus of the petiole (where it meets the leaf). Leaf width was measured as the distance across the widest portion of leaf surface, perpendicular to the midrib. Petiole length was measured from leaf axil to distal terminus of the petiole using a ruler. The distal terminus of the petiole was considered to be located at the line formed by connecting the points where the right and left side leaf margins coincide with the petiole. Petiole width was measured using a caliper near the distal terminus of the petiole at the point where the petiole first assumes a uniform width. Width was measured perpendicular to the leaf midrib. The petiole is curved at the terminus where it fuses with leaf margins. Measurements were not taken in the curved part, but further towards the leaf axil. Leaf thickness was measured with a caliper to one side of

**Table 1** Location information of accession collecting sites

ID	PI No.	Latitude	Longitude	Elevation (m)	Location notes
1	504266	41.38889	9.165556	4	Bonifacio, Ajaccio, Corsica
2	504269	41.51667	9.216667	1	Porto Vecchio, Ajaccio, Corsica
3	504279	41.91611	8.739167	1	Ajaccio, Corsica
4	504181	42.4	9.516667	10	Prunete, Bastia, Corsica
5	504277	42.63278	8.942222	5	I'lle Rousse, Bastia, Corsica
6	504273	42.70222	9.450833	10	Grigione, Bastia, Corsica
7	540562	43.46667	4.333333	0	Mas de Cabassale, Cumarque County
8	540578	45.3	-0.783333	2	LaMarechale, Gironde County
9	540582	45.8	-1.15	5	la Tremblade les Brandes, Charente Maritime County
10	540592	46.24417	-1.561111	1	Ils-De-Re' Plane-des-Baleines, Charente Maritime County
11	540595	46.3	-1.016667	3	Marans, Charente Maritime County
12	540599	46.63333	-1.866667	8	Bretgnolles-Sur-Mer, Vendee County
13	540602	47.01667	-1.966667	1	Bourgnuf-En-Retz, Loire-Atlantique County
14	540606	47.23333	-2.15	2	St-Brevin-Les-Pins Minden, Loire-Atlantique County
15	540609	47.66667	-3.166667	1	Lorient, Morbihan County
16	540692	47.76639	-3.548611	1	Le Pouldu, Santec, Finistere County
17	540613	47.93333	-4.390833	3	Quimper, Finistere County
18	540618	48.51667	-4.75	1	St. Renan, Finistere County
19	540637	48.61667	-2.033333	1	LaGouonnais, Ille-Et-Vilaine County
20	540640	48.63333	-1.483333	1	Le Mont-St-Michel, Manche County
21	540641	48.65	-1.4	1	Avranches, Manche County
22	540690	48.78278	-3.0575	1	Paimpol, Cotes-Du-Nord County
23	540645	49	-1.55	1	Regneville, Manche County
24	540656	49.28333	-0.133333	1	Cabourg, Calvados County
25	540647	49.33333	-1.7	1	Portbail, Manche County
26	540651	49.58333	-1.266667	2	St. Vaast La-Hougue, Manche County
27	540661	50.01667	1.333333	2	LeTreport, Seine-Maritime County
28	540665	50.76667	1.616667	2	Wimereux, Pas-De-Calais County

Latitude and longitude given in decimal, elevation given in meters

the midrib, and not on top of any major lateral veins, near the widest portion of the leaf. To remove correlations between trait values, length and width measurements of both the leaf and the petiole were reduced to length/width ratios.

## Data analysis

### *Molecular marker data*

Descriptive statistics, including expected and observed heterozygosities, differentiation between accessions

(overall and hierarchical  $F_{st}$ ) and diversity within accessions (as measured by the absolute number of polymorphic alleles) and a sample adjusted metric of allelic richness useful for group comparisons (El Mousadik and Petit 1996), were estimated from genotypic data using the software package GDA (Lewis and Zaykin 2001) and FSTAT (Goudet 1995; Lewis and Zaykin 2001). Distances between individual genotypes were calculated as a proportion of shared alleles using DARwin software (Perrier and Jacquemoud-Collet 2006). The neighbor-joining tree using these distances was produced within DARwin. Inter-individual distances were rendered as an unrooted

**Table 2** Primer information associated with each of the 13 SSR loci used in the study

Primer ID <sup>a</sup>	Range (bp)		Primer sequence	Linkage group
509a <sup>5</sup>		FORWARD	5'-TGC TCT CAT CAT CTT CTC CAA TAG-3'	
		REVERSE	5'-ATA TTT TTA GTG AAT TTA GAA AG-3'	
FDSB1001 <sup>3</sup>	308–348	FORWARD	5'-ATC TTA TGC TGC CAT GAC CA-3'	
		REVERSE	5'-ACT TCA ACC ACT ATC ACA AAG TGA G	
FDSB1002 <sup>2,4</sup>	143–177	FORWARD	5'-CCT TAA ACC TAA AAA CGC CAG C-3'	4
		REVERSE	5'-GAA AAC GGA GTT CAG TCA GGG A-3'	
FDSB1005 <sup>3</sup>	273–285	FORWARD	5'-AGC TTA GCA TGC TCT TTC TGG TC-3'	
		REVERSE	5'-AAC CTG AGG AGA AGG TGG ATT TG-3'	
FDSB1023 <sup>4</sup>	224–262	FORWARD	5'-GTA GCT AGT TCA GCA ATC TTC GC-3'	4
		REVERSE	5'-TCT CTC TCC CCC TAA AAG TTC A-3'	
FDSB1026 <sup>3</sup>	173–237	FORWARD	5'-ATC AGC AGG AGA ACC CTG AAA TAC T-3'	
		REVERSE	5'-TCC CTT GTG TTC CTC TAG CTT CTT TA-3'	
FDSB1027 <sup>4</sup>	185–221	FORWARD	5'-GCT GGA TGC TGA CAA CTA TGA AAC-3'	3
		REVERSE	5'-CAG GCA TGA GTA GCA TGA ACT AAA G-3'	
GAA1 <sup>2</sup>	173–200	FORWARD	5'-TGG ATG TTG TAC TAA AGC CTC A-3'	
		REVERSE	5'-TCC TAC CAA AAT GCT GCT TC-3'	
GCC1 <sup>2</sup>	97–106	FORWARD	5'-TAG-ACC AAA ACC AGA GCA GC-3'	
		REVERSE	5'-TGC TCT CAT TTC GTA TGC AC-3'	
GTT1 <sup>2,4</sup>	108–126	FORWARD	5'-CAA AAG CTC CCT AGG CTT-3'	6
		REVERSE	5'-ACT AGC TCG CAG AGT AAT CG-3'	
SB09 <sup>1</sup>	123–141	FORWARD	5'-TGC ATA AAA CCC CCA ACA AT-3'	
		REVERSE	5'-AGG GCA ACT TTG TTT TGT GG-3'	
SB11 <sup>1</sup>	161–175	FORWARD	5'-CGA GGG GTA AAA CCA GAC AA-3'	
		REVERSE	5'-GGT TCT GAA ATT TGG GGG TT-3'	
SB13 <sup>1</sup>	93–141	FORWARD	5'-ACA GCA AGA TCA GAG CCG TT-3'	
		REVERSE	5'-TGG ACC CAC CAT TTA CAT CA-3'	

<sup>a</sup> Superscript refers to citation for microsatellites: <sup>1</sup> Richards et al. (2004); <sup>2</sup> Viard et al. (2002); <sup>3</sup> V. Laurent, personal communication; <sup>4</sup> McGrath et al. (2007), and <sup>5</sup> Reeves et al. (2012)

dendrogram using FigTree v1.3.1 (Rambaut 2006; Rambaut and Drummond 2009).

Population structure was evaluated with two complementary Bayesian Markov Chain Monte Carlo (MCMC) methods either using the whole data set of 28 accessions or a subset containing 21 accessions distributed along the French Atlantic coast of France. First, the approach of Huelsenbeck and Andolfatto (2007) was implemented in the software STRUCTURAMA and used to infer the number of genetic clusters (K) from the genotypic data. K was treated as a random variable with a Dirichlet process prior. The concentration parameter was set to 2. Each Markov chain was run for  $1 \times 10^6$  generations. The first half of the run was treated as the burn in period (intended to allow the chain to reach equilibrium), after which the chain was

sampled every 500 generations. A total of 100 independent runs was conducted, each of which provided an estimate of K as the number of subpopulations with the highest posterior probability. The optimal K value was inferred as the most common K value estimated across the independent runs. Second, the software STRUCTURE v2.2 (Pritchard et al. 2000) was used to estimate admixture as the fractional assignment of a genotype among the previously defined K clusters. The largest fractional assignment is the membership coefficient for the assigned cluster. The burn in period was set to  $5 \times 10^5$  generations, after which the chain was sampled for  $1 \times 10^5$  generations. Fifteen replicate runs were conducted. Label switching and the possibility of multimodality among independent runs was corrected using CLUMPP (Jakobsson and Rosenberg

2007) implemented in Structure Harvester (Evanno et al. 2005). The average membership coefficients calculated from aligned assignment matrices were retained as a quantitative estimate of population structure and admixture among the accessions.

### Phenotypic data

The morphological trait measurements (leaf length/width ratio, petiole length/width ratio and leaf thickness) were summarized with descriptive statistics, and analyzed with regression and multivariate analysis using the statistical software JMP v10.0 (SAS Institute, Cary, NC). The morphological traits was analyzed independently as a two-way ANOVA with block and accession\_ID as the main effects. In addition, we analyzed the relationship of trait value and latitude. Each trait was examined independently using a linear fit of latitude as X variable and trait value as the Y. Block was used as a covariate.

To examine the relationship between diversity estimates based on genetic versus morphological data we used Canonical Discriminant Analysis. The goal was to assess if a multivariate phenotype could be used to predict its categorical group identity (population of origin or its assigned genetic cluster from the SSR analysis). For this analysis we used linear discriminant analysis with prior probabilities proportional to their occurrence. Significance was examined graphically with a bi-plot of the first two canonical variables showing the points, mean value of the discriminant score for the group and an ellipse that corresponds to the 95 % confidence interval of the mean. In addition a Wilks Lambda test was used to test the significance of the discriminant function using approximate F-statistics and p-values.

Estimations of migration rates among genetic clusters within the data set were conducted with the software BA v3.0 (Wilson and Rannala 2003). This Bayesian method is used to identify rates of recent immigration from multi-locus data. Most significantly this approach relaxes a few key assumptions typically made in assignments tests or indirect gene flow methods, such as letting genotype frequencies deviate from Hardy–Weinberg equilibrium. To estimate the posterior probabilities of migration, the MCMC chain was run  $2 \times 10^6$  iterations. The first half of the run was used as a burn-in period after which the samples were collected every 2,000 iterations. Upon the

author's recommendation, we adjusted three of the algorithms mixing parameters and performed 10 independent runs to look for acceptance rates of the results and consistency of convergence.

## Results

### Molecular analysis

Each of the loci used in this study displayed a high level of variability with PIC cores ranging from 0.91 to 0.3 (Table 3). This is reflected in the number of alleles per locus and the number of observed genotypes within the entire data set. Data filtering (used to limit missing data to a maximum of one locus per multilocus genotype) reduced the final sample size of the study from 336 (12 individuals each for 28 accessions) to 277 individuals (Table 4).

Patterns of diversity and differentiation across the sample accessions revealed both clinal gradients and admixture. The observed heterozygosity was high across the set of accessions (mean of 0.48). The mean inbreeding coefficient ( $f$ ) was (0.13), and significantly greater than zero. Variation for these two parameters showed a gradient of values across latitudes within both the entire set of 28 accessions and within only the 21 French Atlantic accessions. Inbreeding increased

**Table 3** Descriptive statistics for each SSR locus

Marker	N genotypes	N alleles	He	Allelic Richness*	PIC
509a	22	8	0.71	3.72	0.67
FDSB1001	46	17	0.5	3.09	0.5
FDSB1002	56	16	0.85	5.14	0.83
FDSB1005	16	7	0.71	3.37	0.65
FDSB1023	31	12	0.79	4.22	0.75
FDSB1026	124	31	0.92	6.32	0.91
FDSB1027	52	17	0.67	3.88	0.66
GAA1	10	6	0.33	2.09	0.3
GCC1	5	3	0.46	2.02	0.36
GTT1	14	6	0.6	2.9	0.56
SB09	14	7	0.62	2.88	0.54
SB11	20	8	0.57	3.18	0.54
SB13	17	9	0.66	3.18	0.61

He is expected heterozygosity, PIC value is polymorphic information content

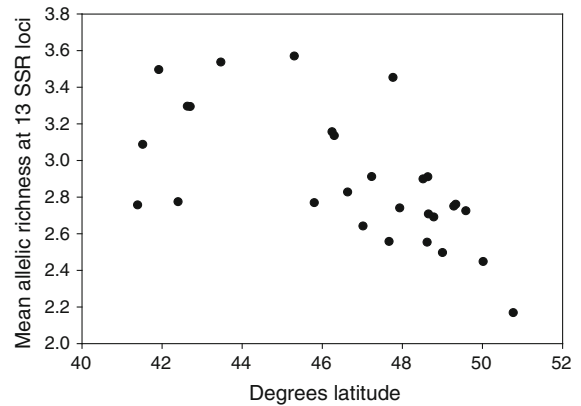


**Table 4** Descriptive statistics for each accession used in the study

ID	PI no.	N samples	Weighted N	He	Ho	f
1	504266	12	11.69	0.52	0.52	0
2	504269	8	7.54	0.5	0.51	-0.02
3	504279	12	11.46	0.67	0.63	0.06
4	504181	12	11.23	0.52	0.47	0.1
5	504277	12	11.31	0.61	0.58	0.05
6	504273	8	7.69	0.59	0.49	0.18
7	540562	12	11.15	0.67	0.54	0.2
8	540578	11	10.85	0.69	0.64	0.07
9	540582	8	6.85	0.55	0.44	0.21
10	540592	12	11.38	0.6	0.47	0.23
11	540595	8	7.31	0.62	0.53	0.15
12	540599	8	7.77	0.54	0.5	0.08
13	540602	12	11.62	0.5	0.41	0.18
14	540606	11	10.62	0.54	0.46	0.16
15	540609	8	7.62	0.46	0.43	0.09
16	540692	8	7.77	0.67	0.69	-0.04
17	540613	10	9.15	0.51	0.47	0.08
18	540618	8	7.92	0.58	0.47	0.21
19	540637	8	7.77	0.44	0.37	0.17
20	540640	12	11.08	0.54	0.45	0.17
21	540641	12	10.85	0.56	0.51	0.08
22	540690	7	6.77	0.47	0.47	0.01
23	540645	12	11.31	0.49	0.41	0.17
24	540656	8	7.69	0.61	0.54	0.11
25	540647	11	10.46	0.47	0.33	0.32
26	540651	8	7.54	0.56	0.36	0.37
27	540661	11	10.77	0.49	0.38	0.23
28	540665	8	7.85	0.4	0.29	0.3

Weighted N takes into account missing data in genotyping scores, He is expected heterozygosity, Ho is observed heterozygosity and f is a inbreeding coefficient

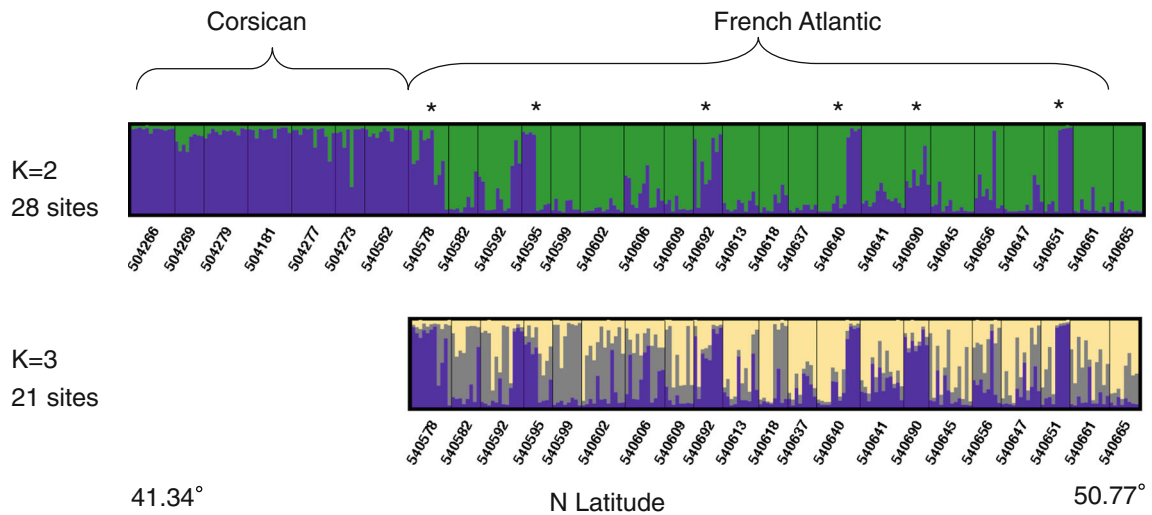
northward while heterozygosity increased southward which support the idea of northward colonization from species center of diversity in the Mediterranean basin. Each regression was significant at the  $p < 0.05$  level with  $R^2$  of 0.30 for observed heterozygosity and an  $R^2$  of 0.23 for inbreeding coefficient. In addition, we examined the pattern of diversity as measured by allelic richness, which controls for variation in sample sizes across the French Atlantic sampling region. This regression is significant at the  $p < 0.001$  level with an  $R^2$  of 0.45 and shows a decrease in allelic richness with increased latitude (Fig. 2). By contrast, isolation by distance was not supported. Differentiation measured



**Fig. 2** Bi-plot of allelic diversity in SSR loci as a function of sample site latitude. One way ANOVA is significant at  $p = 0.0003$ . Coefficient of determination ( $R^2$ ) was 0.39

as average  $F_{st}$  among all accessions was 0.14 with a 95 % confidence interval of (0.17–0.11). A Mantel test between the French Atlantic accessions revealed no significant correlation between pair-wise differentiation ( $F_{st}$ ) and geographic distance (data not shown).

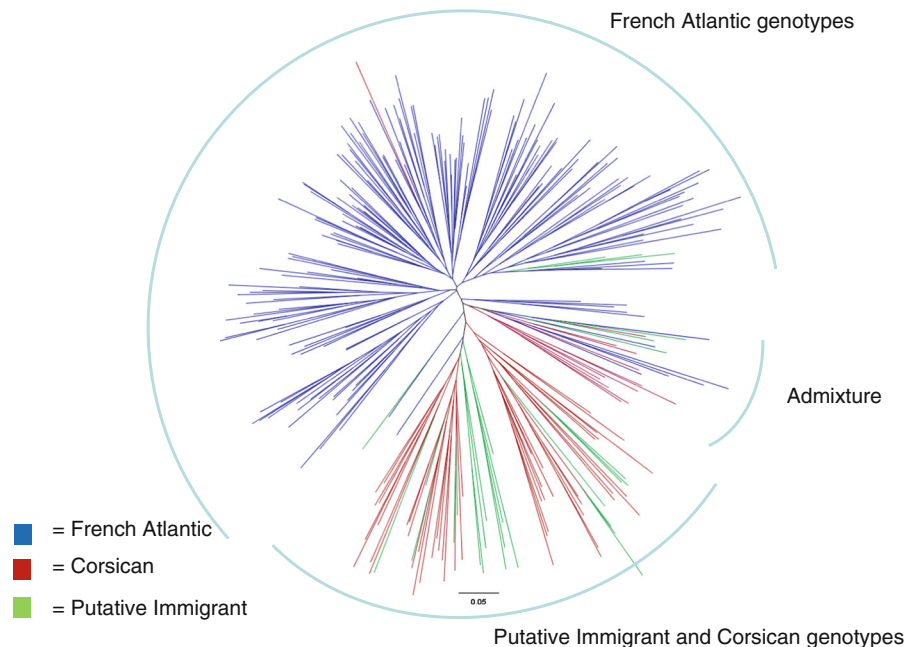
Clustering the genotypic data using Bayesian model-based methods revealed a strong genetic discontinuity between the Mediterranean and French Atlantic accessions. The mode value for K was found to be two for the entire data set. The average posterior probability was 0.98. Among 100 independent runs K = 2 occurred 96 times. The mode value for K among the 21 French Atlantic accessions was three with an average posterior probability of 0.82 and occurred in 88 % of the independent runs. Among all the 28 accessions, there is a strong genetic discontinuity between the Mediterranean and the French Atlantic samples (Fig. 3a). However the pattern of discontinuity is not precisely defined by these two regions. Genotypes along the French Atlantic coast were often assigned to the genetic cluster associated with the Mediterranean accessions. The assignment pattern observed in the whole data set also reappears when the analysis is run at K = 3 for just the French Atlantic accessions (Fig. 3b). From these observations of genetic diversity we could identify three distinctive genetic groups: a Mediterranean type (consisting of the Corsican samples), an French Atlantic group, and a set of genotypes found among accessions along the French Atlantic that assign strongly (membership coefficient above 0.8) to the Mediterranean type,



**Fig. 3** Genetic clustering of SSR genotypes grouped by accession and ordered from left to right from southern most to northern most. *Top panel* shows structure clustering for all 28 accessions. The full dataset can be stably partitioned into a

Corsican and French Atlantic genetic groups. *Bottom panel* partitions only the 21 French Atlantic accessions into three stable lineages. *Starred accessions* indicated the presence of putative immigrants

**Fig. 4** Unrooted neighbor-joining tree using inter-individual genetic distances



which we classify as a putative French Atlantic immigrant group. Admixture among these genotypes appears almost bi modal, with genotypes showing little intermediacy in membership coefficient. A neighbor-joining tree constructed with inter-individual distances illustrates the relationship among these 3

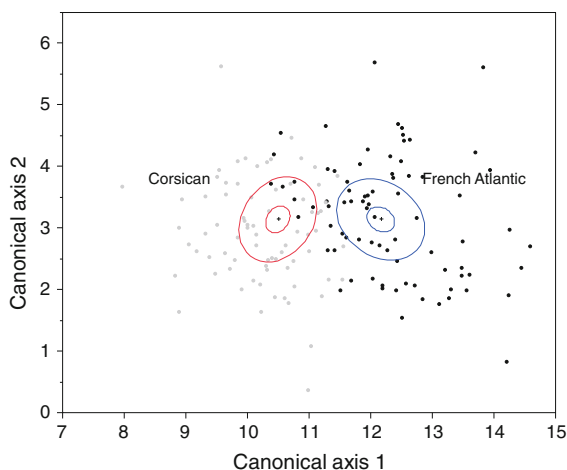
groups (Fig. 4). The tree structure shows that putative French Atlantic immigrant genotypes cluster predominantly with the Mediterranean genotypes rather than French Atlantic genotypes. In addition an admixed group that contained all three types was apparent (marked admixture in Fig. 4).



## Phenotypic analysis

Phenotypic foliar traits were significantly different among accessions in two-way ANOVA using block and accession\_ID as the main effects and either leaf/length ratio, petiole length/width ratio or leaf thickness as the response variable. Leaf length/width ratio, petiole length/width ratio and leaf thickness were significant at the 0.001, 0.05 and 0.001 levels respectively. The linear regression of these three traits on latitude resulted in significance for leaf length/width ratio ( $y = 0.75 + 0.02x$ ,  $p < 0.001$ ) and leaf thickness ( $y = -0.32 + 0.02x$ ,  $p < 0.001$ ) but not petiole length/width ratio ( $p = 0.21$ ). Both leaf length/width ratio and leaf thickness increased with increasing latitude but linear model had low predictive power ( $R^2 = 0.05$  and  $0.16$ , respectively).

Combining all three traits into a multivariate discriminant analysis of all 277 individuals using marker-derived group assignments of  $K = 2$  (Fig. 5), resulted in a discriminant function that could correctly predict group membership 75 % of the time (Wilks Lambda test was significant at  $p < 0.001$ ). Grouping the discriminant analysis by  $K = 3$  groups based on genetic assignment (Corsican, French Atlantic and Putative Immigrant), resulted in a clear separation of French Atlantic genotypes from the Corsican. Corsican and



**Fig. 5** Canonical bi-plot shows the points and multivariate means using phenotypic traits from 277 individual plants grown in a common garden. *Small ellipses* correspond to the 95 % confidence interval for the mean in each category. *Larger ellipses* shows areas that contain roughly 50 % of the points for that group

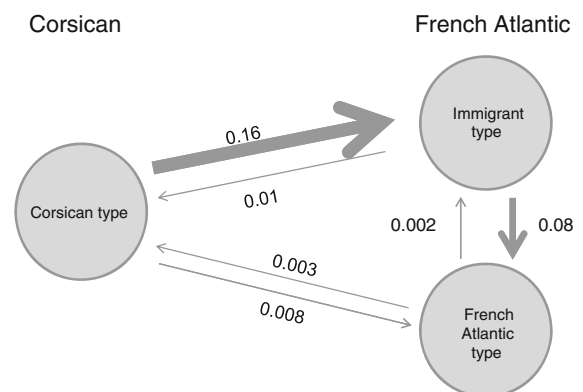
Putative Immigrant groups could not be statistically separated (data not shown).

## Estimation of migration among groups

Migration rates were estimated among three classes of genotypes (Mediterranean, French Atlantic, and putative French Atlantic immigrant) using a Bayesian estimator (Fig. 6). Rates of migration were notably heterogeneous among the three groups with significant directional asymmetry. The rates of migration were quite low between the Mediterranean and French Atlantic groups in both directions. The rate of migration from the Mediterranean group to the putative French Atlantic immigrant group was 20 times higher than the migration rate from the Mediterranean group to the French Atlantic group. The migration between the French Atlantic and the French Atlantic immigrant group was also asymmetrical with migration 40-fold higher from the French Atlantic immigrant group to the French Atlantic group than the reverse.

## Discussion

The genetic structure of plant populations is shaped by both landscape features and demographic processes that involve both historical and contemporary gene flow. Often the pattern is estimated indirectly at a coarse grained, population level (Schwartz and McKelvey 2009). Advances in analytical methods



**Fig. 6** Schematic diagram of directional gene flow rates estimated using BA3 among Corsican, French Atlantic and putative immigrant genotypes as identified in Structure analysis. *Arrow thickness* roughly approximates gene flow rates that are given as values

make it possible to estimate the genetic origin of individual immigrant genotypes (Waples and Gaggiotti 2006). These methods do not look just for broad signatures of differentiation at the population level but are able to quantify the effect of admixture on individual genotypes and may better describe the biological reality of the population structure within a species.

Distributions of many species in Europe have been impacted by cycles of glaciation in that region of the world (Hewitt 1999, 2000; Schmitt 2007). This pattern of phylogeographic distribution is also observed in *B. maritima* through variation in the chloroplast haplotype data [Villain (2007, 2009; reviewed by Biancardi et al. 2012, Chap. 3.22)]. Differences between French Atlantic and Mediterranean populations of sea beet have been noted by others including Doney (1992) and Letschert (1993), who also found greater diversity among the Mediterranean populations using allozyme analyses. Sea beet has the Mediterranean as its center of diversity and, most likely, center of domestication (Doney 1992; Viard et al. 2004; Fievet et al. 2007, Ford-Lloyd et al. 1975; Biancardi et al. 2012). Although Letschert and Frese (1993) attribute at least part of the greater variation in Mediterranean populations of sea beet to the ‘dynamic and variable habitat’, it is likely that the associated drift effects from colonization of the North French Atlantic coast from Mediterranean refugia after the last ice-age has allowed less time for these populations to develop the diversity seen in the longer colonized habitats in the Mediterranean regions.

As a consequence of this biogeographic history several authors have described a continuous latitudinal cline in life history traits that occurs among populations along the French Atlantic coast of France (Van Dijk et al. 1997; Boudry et al. 2002; Hautekèete et al. 2002a; Wagmann et al. 2012). In these studies it was demonstrated that there is a complex north–south cline in flowering time across the French Atlantic sampling region. These data suggest that flowering time is a consequence of selection on photoperiod sensitivity and vernalization requirement across this region (Reeves et al. 2012). The distribution of these traits is thought to be influenced by gene flow and demographic dynamics at both metapopulation and regional scales (Van Dijk 2009).

Our results support a phylogeographic pattern that separates the French Atlantic accessions from our

reference Corsican accessions. Viewed at the level of sampled accessions, the allelic richness and heterozygosity decrease while inbreeding levels increase northward in a pattern consistent with glacial oscillations and range expansion and contraction. The main genetic discontinuity identified in the Bayesian analysis stably clusters the Corsican accessions from the rest of the French Atlantic accessions. This structure is also supported by phenotypic discriminant analysis using foliar traits.

The broad genetic and phenotypic discontinuity appears to be overlaid with a more complex pattern of recent gene flow. While Bayesian clustering separates Corsican from French Atlantic genotypes, the data also show a recurrent pattern of Corsican types up the French Atlantic coast. The pattern might be expected as gene flow among these coastal sampling sites creates a gradient of admixture; however the observed pattern shows a discrete and localized grouping of Corsican genotypes. The Bayesian analysis showed notable bi-modal membership coefficients where individual genotypes are either strongly and unambiguously assigned to the Corsican or the French Atlantic cluster including the putative immigrants. Further analysis of these putative immigrants shows that they are phenotypically distinguishable from the more common French Atlantic phenotypes and indistinguishable from the Corsican types. Estimate of reciprocal gene flow among putative immigrants and French Atlantic genotypes are highly asymmetrical. Taken together, the lack of intermediacy in the genotypic assignments, the phenotypic differentiation and the asymmetry in estimated gene flow suggest that immigrants identified in this study may be of recent origin and could represent the result of an ongoing dispersal dynamic. While the sampling design does not allow us to determine the specific origin of the putative immigrants (sources outside the sampling accessions cannot be excluded), our analysis supports the hypothesis that what once considered a gradual cline is actually an admixed mosaic of genotypes.

Statistical tests that describe the geographic structure of intra specific variation are essential to many types of genetic resources research. Accurate estimation of genetic discontinuities and regions of high diversity are important for locating populations that may contain novel adaptations. Although isolation by distance has been demonstrated in many plant species, geography cannot always be a surrogate for genetic

differentiation: validation is necessary (Wilson and Rannala 2003). This work shows that large scale patterns of genetic structure that might have built over long periods of time are subject to contemporary gene flow dynamics that may create mosaic distributions of genetic diversity that are more challenging to interpret in a strictly spatial context. Accounting for these processes will improve the way we describe the scale of adaptive differentiation and may help improve the structure and utilization of genetic resources collections.

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